

# **Product Data Sheet**

# MDA-MB-468 A-FLX™ FFPE Cell Pellet

## GENERAL INFORMATION

Product Name:	MDA-MB-468 A-FLX™ FFPE Cell Pellet	
Reference Number:	3010-0410	Block
	3010-0420	Slide (4µm)
	3010-0430	FFPE scroll (20µm)
Date of Manufacture:	See product label	
Lot Number:	See product label	
Intended Use:	For research use only	

### DESCRIPTION

Cell Line:	MDA-MB-468	
Tissue of Origin:	Breast	
Culturing Condition:	DMEM supplemented with 10% FBS at 37°C with 5% $\ensuremath{\text{CO}_2}$	
Fixation Condition:	10% neutral buffered formalin (NBF) for 24 hours at 24-27°C	
Product Format:		
Block:	Paraffin embedded block.	
	Pellet diameter: 5mm	
	Sections: 300+ sections at 4 $\mu$ m	
Slide:	One unstained section mounted on Superfrost™ Plus slide. Section thickness: 4µm	
FFPE Scroll:	One FFPE section in DNase/RNase free tube. Section thickness: 20µm	

### **SCHEMATICS**

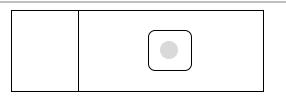


Illustration of an FFPE slide

# STORAGE CONDITION

Block & Slide: 2-8 °C with desiccation (Recommended). For extended shelf life, store samples at -15 °C to -5 °C with desiccation.

# \*RECOMMENDED STABILITY RE-TESTING SCHEDULE:

Block	5 years
Scroll	1 year
Slide	1 year

\*FFPE sample stability is not universally established and is biomarker and assay dependent. More frequent re-testing may be needed for certain labile biomolecules.

# SAFETY AND PRECAUTIONS

This product does not contain hazardous material. Wear appropriate personal protective equipment (PPE) when handling reagents and biological specimens.

# **RECOMMENDED PROCEDURES**

### Staining using FFPE slides:

- 1. Bake slides at 60°C for 30-60 min.
- 2. Deparaffinize 3 times in Xylene or Xylene substitute for 5 min each time.
- 3. Rinse 2 times in 100% ethanol for 1 min each.
- 4. Rehydrate in ethanol series (95% 1 min, 70% 1 min, distilled  $H_2O$  2 times for 1 min each).
- 5. Proceed to staining protocol.

### Biomolecule extraction from FFPE scrolls:

- Add 1ml Xylene or Xylene substitute to each tube containing FFPE scrolls and vortex for 30 sec.
- 2. Centrifuge at full speed for 2 min at room temperature. Remove supernatant without disturbing the pellet.
- 3. Repeat step 1 and 2 for a total of 3 times
- 4. Add 1ml 100% ethanol and mix by vortexing.
- 5. Centrifuge at full speed for 2 min at room temperature. Remove supernatant without disturbing the pellet.
- 6. Repeat step 4 and 5.
- 7. Carefully remove any residual ethanol in the tube without disturbing the pellet.
- 8. Open the tube and dry at room temperature or 37°C for 30min. Ensure that ethanol has completely evaporated.
- 9. Proceed to extraction protocol.



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