

# **Product Data Sheet**

# BT549 A-FLX™ FFPE Cell Pellet

#### GENERAL INFORMATION

Product Name: BT549 A-FLX™ FFPE Cell Pellet

Reference Number: 3010-0910 Block

3010-0920 Slide (4µm)

3010-0930 FFPE scroll (20µm)

Date of Manufacture: See product label
Lot Number: See product label
Intended Use: For research use only

# **DESCRIPTION**

Cell Line: BT549
Tissue of Origin: Breast

Culturing Condition: RPMI-1640 supplemented with 10% FBS at 37°C with

5% CO<sub>2</sub>

Fixation Condition: 10% neutral buffered formalin (NBF) for 24 hours at 24-

27°C

Product Format:

Block: Paraffin embedded block.

Pellet diameter: 5mm

Sections: 300+ sections at 4 µm

Slide: One unstained section mounted on Superfrost™ Plus

slide. Section thickness: 4µm

FFPE Scroll: One FFPE section in DNase/RNase free tube. Section

thickness: 20µm

### **SCHEMATICS**

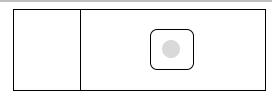


Illustration of an FFPE slide

#### STORAGE CONDITION

Block & Slide: 2-8° C with desiccation (Recommended).

For extended shelf life, store samples at -15 °C to

-5 ° C with desiccation.

### \*RECOMMENDED STABILITY RE-TESTING SCHEDULE:

Block 5 years
Scroll 1 year
Slide 1 year

\*FFPE sample stability is not universally established and is biomarker and assay dependent. More frequent re-testing may be needed for certain labile biomolecules.

#### SAFETY AND PRECAUTIONS

This product does not contain hazardous material. Wear appropriate personal protective equipment (PPE) when handling reagents and biological specimens.

#### RECOMMENDED PROCEDURES

#### Staining using FFPE slides:

- 1. Bake slides at 60°C for 30-60 min.
- 2. Deparaffinize 3 times in Xylene or Xylene substitute for 5 min each time.
- 3. Rinse 2 times in 100% ethanol for 1 min each.
- 4. Rehydrate in ethanol series (95% 1 min, 70% 1 min, distilled  $H_2O$  2 times for 1 min each).
- 5. Proceed to staining protocol.

## Biomolecule extraction from FFPE scrolls:

- Add 1ml Xylene or Xylene substitute to each tube containing FFPE scrolls and vortex for 30 sec.
- 2. Centrifuge at full speed for 2 min at room temperature. Remove supernatant without disturbing the pellet.
- 3. Repeat step 1 and 2 for a total of 3 times
- 4. Add 1ml 100% ethanol and mix by vortexing.
- 5. Centrifuge at full speed for 2 min at room temperature. Remove supernatant without disturbing the pellet.
- 6. Repeat step 4 and 5.
- 7. Carefully remove any residual ethanol in the tube without disturbing the pellet.
- 8. Open the tube and dry at room temperature or 37°C for 30min. Ensure that ethanol has completely evaporated.
- 9. Proceed to extraction protocol.





#### Acepix Biosciences, Inc.

32990 Alvarado-Niles Road, Suite 910 Union City· CA·94587·USA Tel: 1 (888) 866-0580·Web: www.acepixbio.com