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Diagnostik & molekulare Diagnostik



Laborgeräte & Service

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Data Sheet

TCR Activator / PD-L2 Mammalian Expression Kit Catalog #: 60620

Product Description

The recombinant expression vectors are designed to express human engineered T cell receptor (TCR) activator and human PD-L2 (GenBank Accession #NM_025239) in mammalian cells. The transfected cells can be used in conjunction with PD-1/NFAT Reporter/Jurkat T cells (BPS #60535) to study the interactions of PD-1 with PD-L2 ligand in a cellular context and screen for modulators of this signaling pathway.

Background

The binding of Programmed Cell Death Protein 1 (PD-1), a receptor expressed on activated T cells, to its ligands, PD-L1 and PD-L2, negatively regulates immune responses. The PD-1 ligands are found on most cancers, and PD-1:PD-L1/2 interaction inhibits T cell activity and allows cancer cells to escape immune surveillance. The PD-1:PD-L1/2 pathway is also involved in regulating autoimmune responses, making these proteins promising therapeutic targets for a number of cancers, as well as multiple sclerosis, arthritis, lupus, and type I diabetes.

Application

- Screen for activators or inhibitors of PD-1/PD-L2 signaling in a cellular context
- Characterize the biological activity of PD-1 and its interactions with PD-L2

Components

Component	Specification	Amount	Storage
TCR activator + Human PD-L2 (Component A)	Expression vectors constitutively expressing TCR activator and human PD-L2	500 μl (100 ng DNA/μl)	-20°C
TCR activator (Component B)	Expression vector constitutively expressing TCR activator	500 μl (100 ng DNA/μl)	-20°C



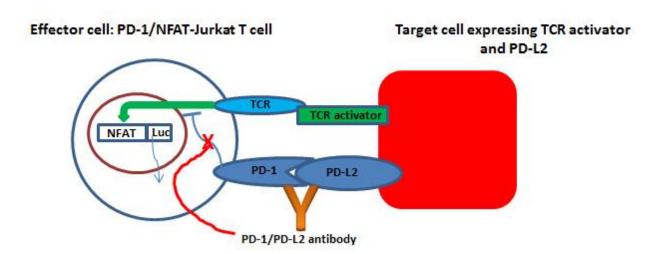
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Functional Validation and Assay Performance

In this assay, PD-1/NFAT Reporter/Jurkat T cells are used as effector cells; HEK293 cells over-expressing PD-L2 and an engineered T cell receptor (TCR) activator are used as target cells. When these two cells are co-cultivated, TCR complexes on effector cells are activated by TCR activator on target cells, resulting in expression of the NFAT luciferase reporter. However, PD-1 and PD-L2 ligation prevents TCR activation and suppresses the NFAT-responsive luciferase activity. This inhibition can be specifically reversed by anti-PD-1 or anti-PD-L2 antibodies. PD-1/PD-L2 neutralizing antibodies block PD-1:PD-L2 interaction and promote T cell activation, resulting in reactivation of the NFAT responsive luciferase reporter.

Assay Principle



Materials Required but Not Supplied

- HEK293 cells and its growth medium or other cell lines
- Transfection reagent for mammalian cell line [We use Lipofectamine™ 2000 (life technologies #11668027). However, other transfection reagents work equally well.]
- PD-1/NFAT Reporter/Jurkat T cells (BPS Bioscience #60535)
- Opti-MEM I Reduced Serum Medium (life technologies #31985-062)
- Assay medium: RPMI1640 + 10% FBS + 1% Penicillin/Streptomycin
- Anti-PD-1 neutralizing antibody (BPS Bioscience #71120)
- 96-well tissue culture-treated white clear-bottom assay plate
- One-Step luciferase assay system: (BPS Bioscience #60690) or other luciferase reagents for measuring firefly luciferase activity
- Luminometer

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Protocol

- 1. One day before transfection, seed HEK293 cells at a density of 35,000 cells per well in $100 \mu l$ of growth medium so that cells will be 90% confluent at the time of transfection.
- 2. Next day, transfect 1 µl of Expression vectors for TCR activator and human PD-L2 (component A) or the control expression vectors for only TCR activator (component B) into cells following the manufacturer's protocol.
- 3. One day after transfection, preincubate the PD-1/NFAT Reporter- Jurkat cells with anti-PD-1 antibody for 30 min prior to adding them to the transfected HEK293 cells.
- 4. After ~16 hours, perform luciferase assay using the ONE-Step luciferase assay system: Add 100 μl of One-Step Luciferase reagent per well and rock at room temperature for ~30 minutes. Measure luminescence using a luminometer. If using luciferase reagents from other vendors, follow the manufacturer's assay protocol.
- Data Analysis: Subtract the average background luminescence (cell-free control wells) from the luminescence reading of all wells.
 The fold induction of NFAT luciferase reporter expression = background-subtracted luminescence of stimulated well / average background-subtracted luminescence of unstimulated control wells.

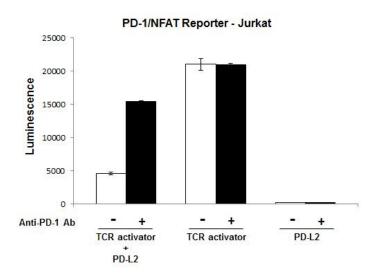
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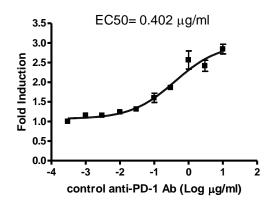
Figure 1. Characterization of biological activity of anti-PD-1 neutralizing antibody in PD-1/PD-L2 cell-based assay using the PD-1/NFAT Reporter-Jurkat cells.

HEK293 cells were transiently transfected with human PD-L2 and an engineered T cell receptor (TCR) activator. The next day, PD-1/NFAT Reporter-Jurkat cells were pre-incubated with anti-PD-1 neutralizing antibody (BPS Cat. #71120) for 30 minutes prior to co-culture with transfected HEK293 cells. After ~16 hours of stimulation, ONE-StepTM Luciferase reagent (BPS Cat. #60690) was added to the cells to measure NFAT activity.

A. Anti-PD-1 neutralizing antibody induced NFAT luciferase reporter activity in PD-1/NFAT Reporter-Jurkat cells co-cultured with HEK293 cells overexpressing PD-L2 and TCR activator.



B. Dose response of anti-PD-1 neutralizing antibody in PD-1/PD-L2 assay in PD-1/NFAT Reporter-Jurkat cells





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Related Products

<u>Product</u>	Cat. #	Size
PD-1 / NFAT - Reporter - Jurkat Recombinant Cell Line	60535	2 vials
NFAT - Reporter - Jurkat Recombinant Cell Line	60621	2 vials
Anti-PD-1 neutralizing antibody	71120	100 µg
ONE-Step [™] Luciferase Assay System	60690-1	10 ml
ONE-Step [™] Luciferase Assay System	60690-2	100 ml
Human PD-1 (CD279), Fc fusion	71106	100 µg
Human PD-1, FLAG-Avi-His-tag	71198	50 µg
Human PD-L1 (CD274), Fc fusion	71104	50 µg
Human PD-L1 (CD274), FLAG-Avi-His tag	71183	50 µg
Human PD-L2 (CD273), Fc fusion	71107	100 µg
Human PD-1, Fc fusion, Biotin-labeled	71109	50 µg
Human PD-L1, Fc fusion, Biotin-labeled	71105	50 µg
Human PD-L2, Fc fusion, Biotin-labeled	71108	50 µg