



# SZABO SCANDIC

Part of Europa Biosite

## Produktinformation



Forschungsprodukte & Biochemikalien



Zellkultur & Verbrauchsmaterial



Diagnostik & molekulare Diagnostik



Laborgeräte & Service

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### Zuschläge

- Mindermengenzuschlag
- Trockeneiszuschlag
- Gefahrgutzuschlag
- Expressversand

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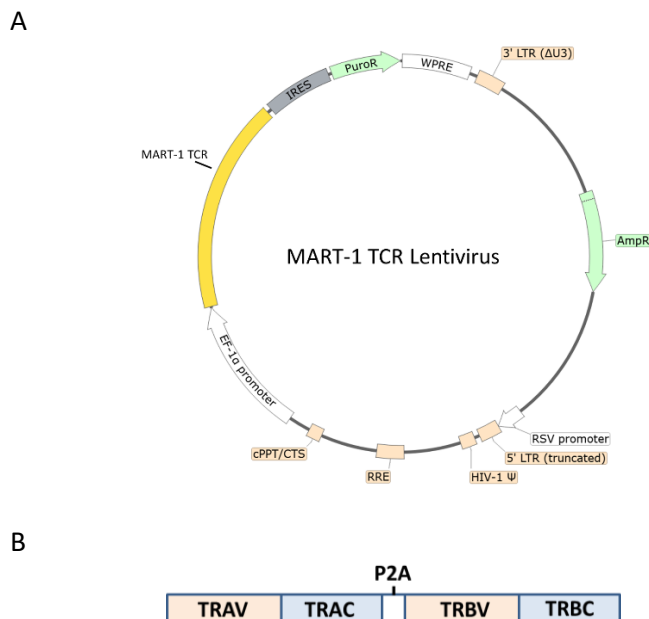
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**Description**

The human MART-1-specific T Cell Receptor lentiviruses (clone DMF5) are replication incompetent, HIV-based, VSV-G-pseudotyped lentiviral particles that are ready to infect almost all types of mammalian cells, including primary and non-dividing cells. These viruses transduce a human TCR (clone DMF5) that specifically recognizes antigen MART-1 (Melanoma-associated antigen recognized by T cells-1, or Melan-A), and in which the TCR  $\alpha$  chain and  $\beta$  chain are linked by P2A (Figure 1). The lentiviruses also transduce a puromycin selection gene.



**Figure 1.** (A) Schematic of the lenti-vector used to generate the MART-1-specific TCR lentivirus and (B) Construct diagram showing expressed components of the MART-1-specific TCR. TRAV and TRAC correspond to the TCR alpha chain variable and constant regions, respectively, whereas TRBV and TRBC correspond to the TCR beta chain variable and constant regions.

**Background**

MART-1 is a differentiation antigen expressed at the surface of melanocytes. A peptide fragment of the protein is found bound to MHC complexes that are recognized by cytotoxic T cells. MART-1 is found in most skin cancers, including melanomas, and is used as a biomarker for diagnostic purposes. Since the expression of the protein is restricted to melanocyte containing tissues (skin and retina), and is not found in other tissues, MART-1 is an attractive target for cancer vaccines and adoptive cell therapy. MART-1 peptide 26-35 is a fragment commonly associated with MHC and recognized by T cell receptors.

**Application**

- Use as a positive control for MART-1-specific TCR evaluation and optimize experimental conditions.
- Generate MART-1-specific TCR stable cell line (puromycin resistant).

**Formulation**

The lentiviruses were produced in HEK293T cells in medium containing 90% DMEM + 10% FBS. Virus particles can be packaged in custom formulations by special request, for an additional fee.

**Titer**

$\geq 2 \times 10^7$  TU/ml. The titer will vary with each lot; the exact value is provided with each shipment.

**Storage**

Lentiviruses are shipped with dry ice. For long term storage, it is recommended to store the virus at -80°C. Avoid repeated freeze-thaw cycles. Titers can drop significantly with each freeze-thaw cycle.

**Biosafety**

The lentiviruses are produced with a SIN (self-inactivation) lentivector which ensures self-inactivation of the lentiviral construct after transduction and integration into the genomic DNA of the target cells. None of the HIV genes (gag, pol, rev) will be expressed in the transduced cells, as they are expressed from packaging plasmids lacking the packing signal. Although the pseudotyped lentiviruses are replication-incompetent, they require the use of a Biosafety Level 2 facility. BPS Bioscience recommends following all local federal, state, and institutional regulations and using all appropriate safety precautions.

**Materials Required but Not Supplied**

These materials are not supplied with this lentivirus but are necessary to follow the designed protocol. BPS Bioscience media, reagents, and luciferase assay systems are all validated and optimized for use with this lentivirus and are highly recommended for best results.

Name	Ordering Information
Thaw Medium 2	<a href="#">BPS Bioscience #60184</a>
Growth Medium 2C	<a href="#">BPS Bioscience #79592</a>
Assay Medium 2D	<a href="#">BPS Bioscience #78755</a>
TCR KO NFAT Luciferase Reporter Jurkat Cell Line	<a href="#">BPS Bioscience #78556</a>
CD8 <sup>+</sup> TCR KO NFAT Luciferase Reporter Jurkat Cell Line	<a href="#">BPS Bioscience #78757</a>
T2 Cell Line	ATCC #CRL-1992
A375 Cells	ATCC #CRL-1619
NY-ESO-1 (157-165) Peptide	<a href="#">BPS Bioscience #78758</a>
MART-1 (26-35) Peptide	<a href="#">BPS Bioscience #78759</a>
MART-1 (Leu26-35, Leu27) Peptide	<a href="#">BPS Bioscience #78760</a>
MART-1 (27-35) Peptide	<a href="#">BPS Bioscience #78761</a>
APC MHC I Dextramer (HLA-A*02:01 ELAGIGILTV)	Immudex #WB02162
PE anti-human $\alpha/\beta$ T Cell Receptor Antibody	BioLegend #306707
ONE-Step™ Luciferase Assay System	<a href="#">BPS Bioscience #60690</a>
Polybrene	Millipore Sigma #TR-1003-G

**Media Formulations**

For best results, the use of BPS Bioscience validated and optimized media is *highly recommended*. Other preparations or formulations of media may result in suboptimal performance.

**Media Required for Maintaining CD8<sup>+</sup> TCR KO NFAT Luciferase Reporter Cell Line**

*Growth Medium 2C (BPS Bioscience #79592):*

RPMI 1640 medium supplemented with 10% FBS, 1% Penicillin/Streptomycin plus 1 mg/ml Geneticin, 100  $\mu$ g/ml Hygromycin B

**Media Required for Maintaining T2 Cells**

*Thaw Medium 2 (BPS Bioscience #60184):*

RPMI 1640 medium supplemented with 10% FBS, 1% Penicillin/Streptomycin

*Media Required for Co-culture Assay**Assay Medium 2D (BPS Bioscience #78755):*

RPMI 1640 medium supplemented with 1% FBS

**Assay Protocol**

The following protocol was used to transduce a Jurkat Cell Line. The transduction conditions (e.g. MOI, concentration of polybrene, time of assay development) should be optimized according to the cell type and the assay requirements. In most cell types, the expression of the reporter gene can be measured approximately 48-72 hours after transduction. For cell types with low transduction efficacy, it may be necessary to select the cells stably expressing the reporter gene with puromycin prior to carrying out the reporter assays.

1. Day 1: Harvest the CD8<sup>+</sup> TCR knockout NFAT Luciferase Reporter Jurkat cells from Growth Medium 2C by centrifugation and resuspend the cells in fresh Thaw Medium 2. Dilute cells to a density of  $2 \times 10^5$ /ml in Thaw Medium 2. Mix 1 ml of the Jurkat cells and MART-1-specific TCR Lentivirus in a 1.5-ml Eppendorf tube at an MOI>10.

Add polybrene to a final concentration of 8 µg/ml. Gently mix and incubate the virus with the Jurkat cells for 20 minutes at room temperature in a tissue culture hood.

2. Centrifuge the virus/cell mix for two hours at 800 x g at 32°C (spinoculation). Add the virus/cell mix from the spinoculation step to one well of a 6-well plate. Add an additional 1.5 ml of Thaw Medium 2 to the well. It is not necessary to remove the virus. Incubate the cells at 37°C with 5% CO<sub>2</sub> for 48-66 hours.

The expression of TCR can be analyzed by flow cytometry. The transduced Jurkat cells are ready for assay development on day 3 or 4. If the transduction efficiency is low, it may be necessary to perform cell selection with puromycin on day 3.

3. For use in the following co-culture assay at Day 4 prepare materials as follows:
  - a) Preparation of Antigen-mimetic Peptides:  
Thaw the MART-1 peptide of interest at room temperature. Dilute the peptide with Assay Medium 2D at a concentration 5-fold higher than the desired final concentration.

*Note: The peptide stock was dissolved in DMSO at a concentration of 1 mM. The final DMSO concentration in the co-culture assay should not be >0.3%.*

- b) Preparation of Antigen Presenting Cells (APCs):  
Harvest T2 cells (APC) from Thaw Medium 2 and resuspend the cells into Assay Medium 2D at a density of  $5 \times 10^5$ /ml. Add 40 µl of T2 cells into each well of a 96-well plate.

Add 20 µl of diluted peptide to the “peptide stimulated” wells. Add 20 µl of Assay Medium 2D to the “unstimulated control” wells (for measuring the basal luciferase activity).

- c) Resuspend Jurkat cells into Assay Medium 2D at a density of  $4 \times 10^5$ /ml.  
Add 40 µl of TCR-transduced CD8<sup>+</sup> TCR knockout NFAT Luciferase Reporter Jurkat cells from Step 3.1 onto the APCs into each well of the 96-well plate containing the APCs.

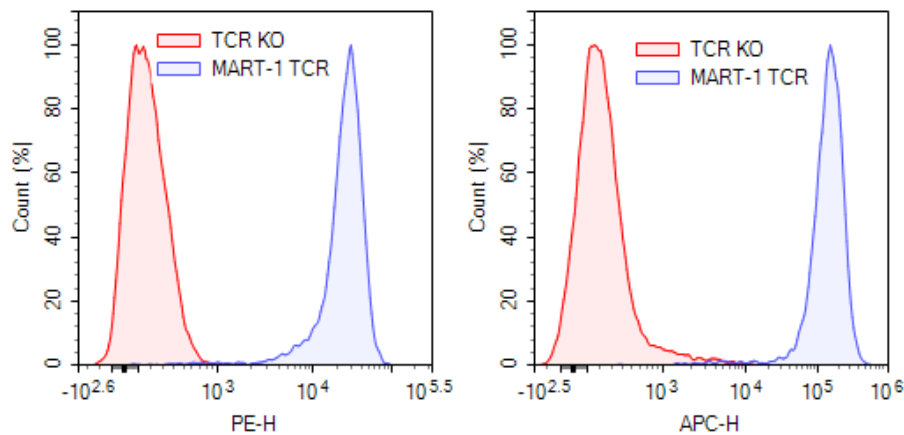
4. Incubate the co-culture plate at 37°C with 5% CO<sub>2</sub> for 5-6 hours or overnight.

5. Prepare the ONE-Step™ Luciferase reagent per recommended protocol. Add 100 µl of ONE-Step™ Luciferase Assay reagent per well. Incubate at room temperature for ~15 to 30 minutes and measure luminescence using a luminometer.

### Notes

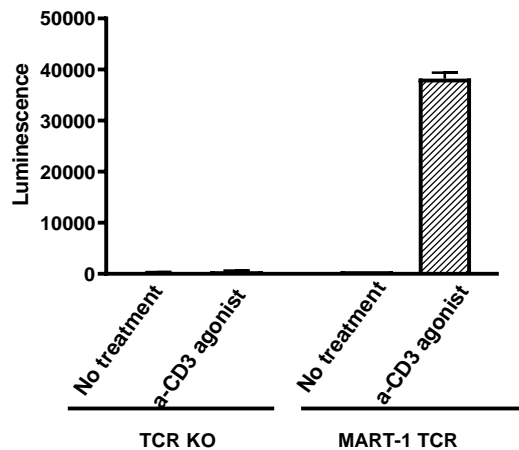
To generate stable TCR-expressing cells, remove the growth medium 48-72 hours after transduction and replace it with fresh growth medium containing the appropriate amount of puromycin for antibiotic selection of transduced cells. For more information on how to determine the concentration of antibiotic required, see our FAQs resources "[what is a kill curve?](#)"

### Validation Data



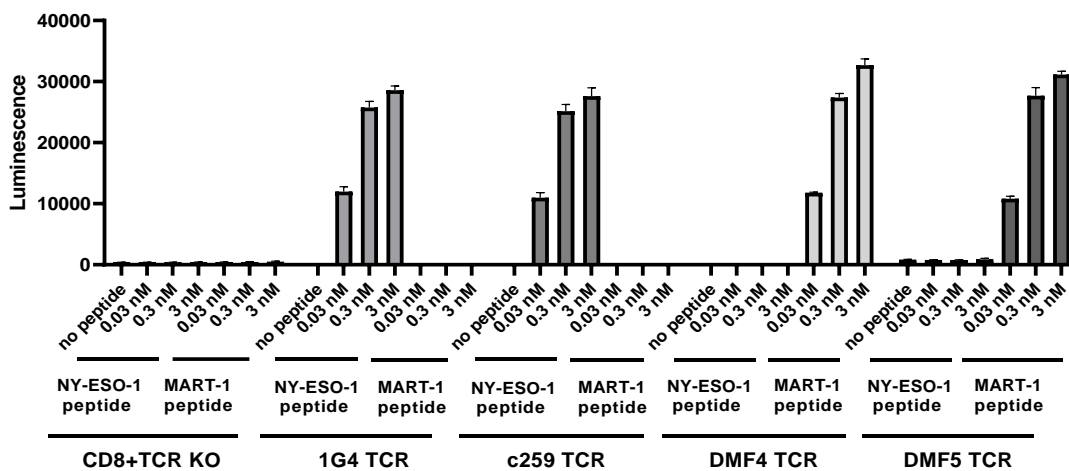
*Figure 2. Expression of MART-1-specific TCR in Jurkat cells transduced with lentiviruses.*

Approximately 100,000 CD8<sup>+</sup> TCR knockout NFAT Luciferase Reporter Jurkat cells (BPS Bioscience #78757) were transduced with MART-1-specific TCR lentivirus (clone DMF5) (BPS Bioscience #78679) via spinoculation at a MOI of 10. After 48 hours of transduction, the cells were transferred into a medium containing 0.5 µg/ml of puromycin. After one week of antibiotic selection, the expression of MART-1-specific TCR (DMF5) was analyzed by flow cytometry. Left: cells were stained with PE conjugated anti-human TCR antibody (Biolegend #306707); right: cells were stained with APC-conjugated MHC-I Dextramer (HLA-A\*02:01 ELAGIGILTV; Immudex#WB02162).



**Figure 3.** Expression of MART-1-specific TCR (clone DMF5) in TCR knockout NFAT Luciferase Reporter Cell Line confers responsiveness to anti-CD3 agonist and induces NFAT-dependent luciferase activity.

Approximately 20,000 TCR knockout NFAT Luciferase Reporter Jurkat cells (BPS Bioscience #78556)/well (96-well plate) were transduced with MART-1-specific TCR lentivirus (clone DMF5) (BPS Bioscience #78679) via spinoculation at a MOI of 10. After 66 hours of transduction, the medium was changed to Thaw Medium 2 (BPS Bioscience #60184). Cells were stimulated by transferring them to a 96-well plate pre-coated with anti-CD3 agonist antibody (BPS Bioscience #71274) at 1 µg/ml for 6 hours. The non-coated wells and the non-transduced cells were run in parallel as controls. Results are shown as raw luminescence readings.



**Figure 4.** T Cell Activation using T2 cells as APC.

CD8<sup>+</sup> TCR knockout NFAT Luciferase Reporter Jurkat cells (BPS Bioscience #78757) were transduced with lentiviruses expressing various TCRs via spinoculation at a MOI of 10. After 66 hours of transduction, the cells were co-cultured with T2 cells (ATCC #CRL-1992) loaded with NY-ESO-1 peptide (BPS Bioscience #78758) or with MART-1 peptide (BPS Bioscience #78760) for 6 hours. The luciferase assay was performed, and the results are shown as raw luminescence readings. Cells transduced with NY-ESO-1-specific TCR (clones 1G4, BPS Bioscience #78765, and c259, BPS Bioscience #78766) can be activated by NY-ESO-1 peptide, but not MART-1 peptide, while cells transduced with MART-1-specific TCR (clones DMF4, BPS Bioscience #78678, and DMF5, BPS Bioscience #78679) can be activated by MART-1 peptide, but not NY-ESO-1 peptide. Un-transduced CD8<sup>+</sup> TCR Knockout NFAT Luciferase reporter cell line, where no TCR is expressed, was run in parallel as a negative control.

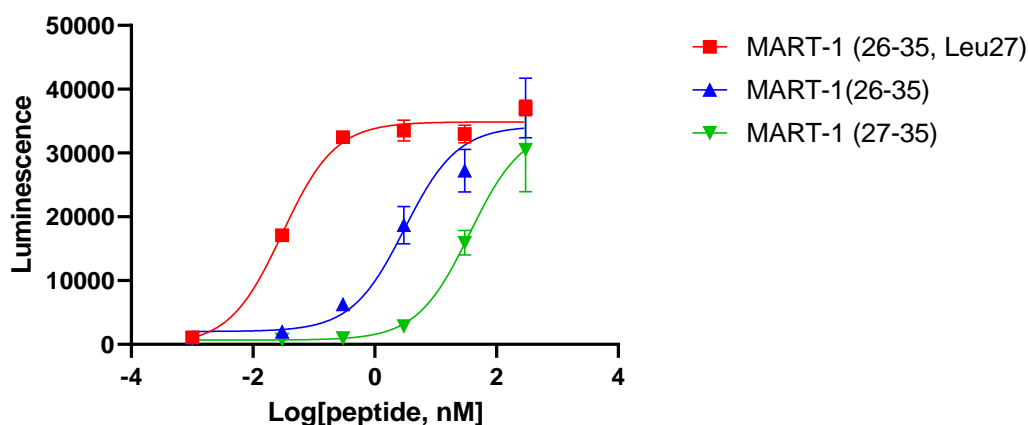


Figure 5. Antigenic Peptide Potency Ranking.

CD8<sup>+</sup> TCR knockout NFAT Luciferase Reporter Jurkat cells (BPS Bioscience #78757) were transduced with lentiviruses expressing MART-1 TCR (Clone DMF5) (BPS Bioscience #78679) via spinoculation at a MOI of 10. After 66 hours of transduction, the cells were co-cultured with different MART-1 peptide loaded T2 cells for 6 hours. The luciferase assay was performed, and the results are shown as raw luminescence readings. The native MART-1 decapeptide (26-35, EAAGIGILTV, BPS Bioscience #78759) is more efficiently recognized by MART-1 TCR (clone DMF5) transduced CD8<sup>+</sup> TCR Knockout NFAT Luciferase Jurkat cells than the MART-1 peptide (27-35, AAGIGILTV, BPS Bioscience #78761), but has lower binding affinity and stability than the MART-1 (26-35, Leu27; ELAGIGILTV, BPS Bioscience #78760) analog.

### Troubleshooting Guide

Visit [bpsbioscience.com/lentivirus-faq](https://bpsbioscience.com/lentivirus-faq) for detailed troubleshooting instructions. For all further questions, please email [support@bpsbioscience.com](mailto:support@bpsbioscience.com).

### References

1. Chodon T. *et al.* Adoptive transfer of MART-2 T-cell receptor transgenic lymphocytes and dendritic cell vaccination in patients with metastatic melanoma. *Clin Cancer Res.* (2014) 20 (9): 2457-65.
2. Rohaan M. *et al.*, MART-1 TCR gene-modified peripheral blood T Cells for the treatment of metastatic melanoma: a phase I/IIa clinical trial. *Immuno-oncol Technol.* (2022) Jun18: 15.
3. Kropp KN. *Et al.*, A bicistronic vector backbone for rapid seamless cloning and chimerization of  $\alpha\beta$ T-cell receptor sequences. *PLOS One* (2020) 15(9): e0238875.

### Related Products

Products	Catalog #	Size
NY-ESO-1-Specific TCR Lentivirus (Clone 1G4)	78675	100 $\mu$ l/2 x 500 $\mu$ l
NY-ESO-1-Specific TCR Lentivirus (Clone c259)	78676	100 $\mu$ l/2 x 500 $\mu$ l
MART-1-Specific TCR Lentivirus (Clone DMF4)	78678	100 $\mu$ l/2 x 500 $\mu$ l
NY-ESO-1 Peptide (157-165)	78758	100 $\mu$ l
MART-1 Peptide (26-35)	78759	100 $\mu$ l
MART-1 Peptide (26-35, Leu 27)	78760	100 $\mu$ l
MART-1 Peptide (27-35)	78761	100 $\mu$ l