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Zuschläge

- Mindermengenzuschlag
- Trockeneiszuschlag
- Gefahrgutzuschlag
- Expressversand

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Description

Notch1dE Lentiviruses are replication incompetent, HIV-based, VSV-G pseudotyped lentiviral particles that are ready to transduce most mammalian cells, including primary and non-dividing cells. These lentiviruses express a truncated human Notch1 construct (Notch1dE) in which the entire extracellular domain was deleted. The lentiviruses also contain a puromycin selection marker (Figure 1). Once expressed in a cell line of interest, Notch1dE is cleaved by γ -secretase, resulting in the constitutively active intracellular domain of Notch (NICD). NICD translocates to the nucleus, binds to the transcription factor CSL (CBF1/RBPJk/Suppressor of Hairless/Lag-1) and activates transcription of Notch1-responsive genes.

When used in combination with CSL (CBF1/RBP-Jk) Luciferase Reporter Lentiviruses (BPS Bioscience #78746), NICD activates the CSL (CBF1/RBPJk/Suppressor of Hairless/Lag-1) responsive elements, inducing luciferase expression.

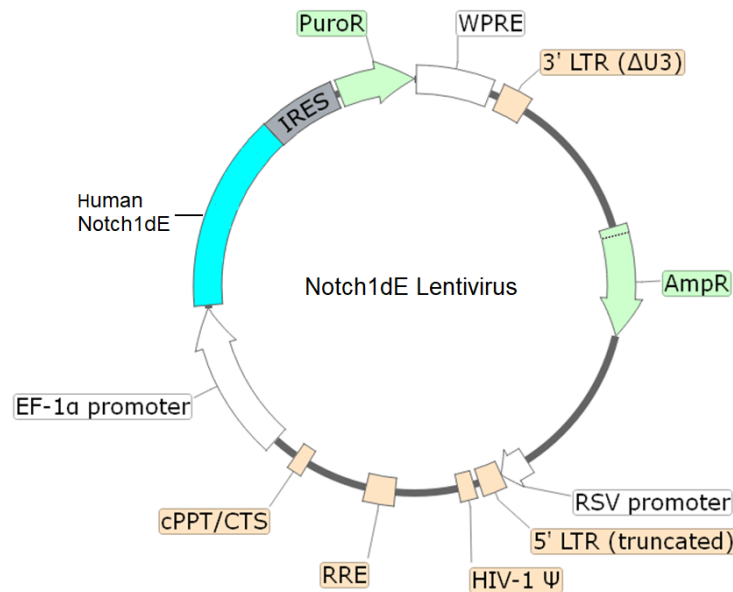


Figure 1. Schematic of the lenti-vector used to generate the Notch1dE lentivirus.

Background

The Notch signaling pathway controls cell fate decisions in vertebrates and invertebrates' tissues and is involved in embryonic development, tissue homeostasis, and regulation of the immune and angiogenic systems. Notch signaling is triggered through the binding of a transmembrane ligand, present in opposing cells, to one of the four existing Notch transmembrane receptors (Notch1/Notch2/Notch3/Notch4). This results in proteolytic cleavage of the Notch receptor, releasing the constitutively active intracellular domain of the Notch receptor (NICD). NICD translocate to the nucleus and associates with the transcription factor CSL (CBF1/RBPJk/Suppressor of Hairless/Lag-1) and coactivator Mastermind to turn on the transcription of Notch-responsive genes. Dysfunction of Notch signaling has severe consequences, including developmental pathologies or cancer (such as T cell acute lymphoblastic leukemia, T-ALL, and urothelial bladder cancer). The use of Notch inhibitors, mainly gamma-secretase inhibitors, as a cancer therapy option and in the regeneration of tissues is ongoing. Further studies will allow us to have a deeper understanding of Notch signaling and will benefit future therapeutic approaches.

Application

- Monitor Notch signaling pathway activity, when used in combination with the CSL (CBF1/RBP-Jk) Luciferase Reporter Lentivirus (BPS Bioscience #78746).
- Generate Notch1dE-expressing cell pools or stable cell lines by puromycin selection.

Formulation

The lentivirus particles were produced in HEK293T cells. They are supplied in cell culture medium containing 90% DMEM + 10% FBS. Virus particles can be packaged in custom formulations by special request, for an additional fee.

Size and Titer

Two vials (500 µl x 2) of lentivirus at a titer $\geq 10^7$ TU/ml. The titer varies with each lot; the exact value is provided with each shipment.

Storage



Lentiviruses are shipped with dry ice. For long-term storage, it is recommended to store the lentiviruses at -80°C. Avoid repeated freeze-thaw cycles. Titers can drop significantly with each freeze-thaw cycle.

Biosafety



The lentiviruses are produced with a SIN (self-inactivation) lentivector which ensures self-inactivation of the lentiviral construct after transduction and after integration into the genomic DNA of the target cells. None of the HIV genes (gag, pol, rev) will be expressed in the transduced cells, as they are expressed from packaging plasmids lacking the packing signal and are not present in the lentivirus particle. Although the pseudotyped lentiviruses are replication-incompetent, they require the use of a Biosafety Level 2 facility. BPS Bioscience recommends following all local federal, state, and institutional regulations and using all appropriate safety precautions.

Materials Required but Not Supplied



These materials are not supplied with this lentivirus but are necessary to follow the protocol described in the “Validation Data” section. Media, reagents, and luciferase assay buffers used at BPS Bioscience are all validated and optimized for use with this lentivirus and are highly recommended for best results.

Name	Ordering Information
HEK293 Cells	ATCC #CRL-1573
Thaw Medium 1	BPS Bioscience #60187
CSL (CBF1/RBP-Jk) Luciferase Reporter Lentivirus	BPS Bioscience #78746
96-well tissue culture, clear-bottom, white plate	Corning #3610
One-Step™ Luciferase Assay System	BPS Bioscience #60690
Luminometer	

Media Formulations

For best results, the use of validated and optimized media from BPS Bioscience is *highly recommended*. Other preparations or formulations of media may result in suboptimal performance.

Media Required for the Proposed Assay

Thaw Medium 1 (BPS Bioscience #60187):

MEM medium supplemented with 10% FBS, 1% non-essential amino acids, 1 mM Na pyruvate, 1% Penicillin/Streptomycin.

Assay Protocol

The following protocol is a general guideline for transducing HEK293 cells using the Notch1dE Lentivirus and CSL (CBF1/RBP-Jk) Luciferase Reporter Lentivirus and it is a general guideline only. The optimal transduction conditions (e.g. MOI, concentration of polybrene, time of assay development) should be optimized according to the cell type and the assay requirements. In most cell types, the expression of the reporter gene can be measured approximately 72 hours after transduction. For cell types with low transduction efficacy, it may be necessary to select the cells expressing the reporter gene with puromycin, creating a cell pool or stable cell line, prior to carrying out the reporter assays.

Day 1:

1. Seed HEK293 cells at a density of 5,000-10,000 cells per well in 90 μ l of Thaw Medium 1 into a white, clear bottom 96-well microplate.
2. To each well, add 2 μ l of CSL (CBF1/RBP-Jk) Luciferase Reporter Lentivirus and 5 μ l of Notch1dE Lentivirus.

Optional: Add polybrene to each well to a final concentration of 5 μ g/ml.

3. Gently swirl the plate to mix.
4. Incubate the plate at 37°C with 5% CO₂ for 24 hours.

Day 3:

1. Add 100 μ l/well of ONE-Step™ Luciferase assay reagent.
2. Incubate the plate at Room Temperature (RT) for ~15 to 30 minutes.
3. Measure luminescence using a luminometer.

Validation Data

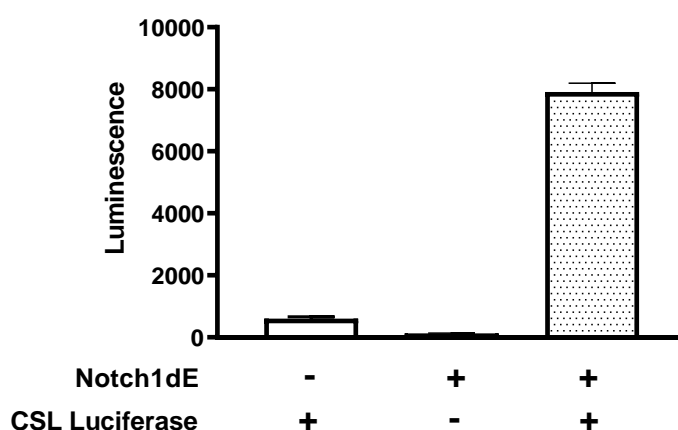


Figure 2. CSL driven luciferase reporter activity in HEK293 cells transduced with CSL (CBF1/RBP-Jk) Luciferase Reporter Lentivirus and/or Notch1dE Lentivirus.

Approximately 10,000 HEK293 cells/well were transduced with 40,000 TU/well of CSL (CBF1/RBP-Jk) Luciferase Reporter Lentivirus and/or 40,000 TU/well of Notch1dE Lentiviruses. After 48 hours

of transduction the luciferase activity was measured with ONE-Step™ Luciferase. Luciferase activity was detected only when both viruses were used. Results are shown as the raw luminescence reading.

Data shown is representative. For lot-specific information, please contact BPS Bioscience, Inc. at support@bpsbioscience.com

Notes

To generate a Notch1dE stable cell line, remove the growth medium 48 hours after transduction and replace it with fresh growth medium containing the appropriate amount of puromycin (as pre-determined from a killing curve, [FAQs \(bpsbioscience.com\)](https://www.bpsbioscience.com/FAQs)) for antibiotic selection of transduced cells, followed by clonal selection.

Reading Luminescence

Luminescence is the emission of light resulting from a chemical reaction. The detection of luminescence requires no wavelength selection because the method used is emission photometry and not emission spectrophotometry. To properly read luminescence, make sure the plate reader is set for LUMINESCENCE mode. Typical integration time is 1 second, delay after plate movement is 100 msec. Do not use a filter when measuring light emission. Typical settings for the Synergy 2 BioTek plate reader: use the “hole” position on the filter wheel; Optics position: Top; Read type: endpoint. Sensitivity may be adjusted based on the luminescence of a control assay.

References

1. Lu F.M., *et al.*, 1996 *Proc. Natl. Acad. Sci. USA* 93(11): 5663-5667.
2. Kanungo J., *et al.*, 2008 *J. Neurochem.* 106: 2236-48.
3. Cao L. *et al.*, 2023 *Blood Adv.* 10.1182/bloodadvances.2023010380

Troubleshooting Guide

Visit [bpsbioscience.com/lentivirus-faq](https://www.bpsbioscience.com/lentivirus-faq) for detailed troubleshooting instructions. For all further questions, please email support@bpsbioscience.com.

Related Products

<i>Products</i>	<i>Catalog #</i>	<i>Size</i>
Negative Control Luciferase Lentivirus	79578	500 µl x 2
CSL (CBF1/RBP-Jk) Luciferase Reporter Lentivirus	78746	500 µl x 2
Notch Signaling Pathway Notch1/CSL Reporter – HEK293 Recombinant Cell Line	60652	2 vials
Notch1 Pathway Reporter Kit (Human)	79503	500 reactions