

Produktinformation



Forschungsprodukte & Biochemikalien



Zellkultur & Verbrauchsmaterial



Diagnostik & molekulare Diagnostik



Laborgeräte & Service

Weitere Information auf den folgenden Seiten! See the following pages for more information!



Lieferung & Zahlungsart

siehe unsere Liefer- und Versandbedingungen

Zuschläge

- Mindermengenzuschlag
- Trockeneiszuschlag
- Gefahrgutzuschlag
- Expressversand

SZABO-SCANDIC HandelsgmbH

Quellenstraße 110, A-1100 Wien

T. +43(0)1 489 3961-0

F. +43(0)1 489 3961-7

mail@szabo-scandic.com

www.szabo-scandic.com

Description

The Chemi-Verse™ STK3 (MST2) Kinase Assay Kit is designed to measure STK3 (serine/threonine-protein kinase 3), also known as MST2, tyrosine kinase activity for screening and profiling applications using ADP-Glo™ as a detection reagent. The assay kit comes in a convenient 96-well format, with enough purified STK3, kinase substrate, ATP, and kinase assay buffer for 100 enzyme reactions.

Background

STK3, also known as serine/threonine-protein kinase 3 or MST2, is involved in the Hippo signaling pathway, a pathway crucial for cell development, proliferation, apoptosis, and stress responses. STK3 is activated by autophosphorylation, which can be triggered by cellular stress signals. Once activated it can phosphorylate LATS1/LATS2 (large tumor suppressor kinase), which in turn inhibits YAP1 (yes-associated protein 1). It was found that kinases such as MAP4K (mitogen-activated protein kinase kinase kinase kinase) and TAOK (thousand and one kinase) function in parallel with STK3/4, depending on the cell type and stress inducer. Inhibited YAP1 cannot translocate to the cell nucleus and activate transcription, being targeted for degradation by the SCF (Skp, cullin, F-box containing) complex. Activated STK3 also phosphorylates FOXO (forkhead box O) factors and leads to transcription of apoptotic genes. Interestingly, STK3 is inhibited by the proto-oncogene c-Raf, and this association is found in many cancers. STK3 can inhibit cancer progression, and it was found at low levels in gastric cancer and hepatocellular carcinoma. Conversely, high levels have also been found in breast cancer and oral squamous cell carcinoma. The development of inhibitors able to disrupt the interaction between c-Raf and STK3 may prove beneficial against some cancer types, but a better understanding of the opposite roles of STK3 in different cancers is still necessary.

Applications

Study enzyme kinetics and screen small molecule inhibitors for drug discovery and high throughput screening (HTS) applications.

Supplied Materials

Catalog #	Name	Amount	Storage
40013	STK3 (MST2), GST-Tag*	2 μg	-80°C
79334	5x Kinase Buffer 1	1.5 ml	-20°C
79686	500 μM ATP	50 μΙ	-20°C
78514	MBP (5 mg/ml)	50 μΙ	-20°C
82545	White 96-well plate	1	Room Temperature

^{*}The concentration of the protein is lot-specific and will be indicated on the tube.

Materials Required but Not Supplied

Name	Ordering Information
ADP-Glo™ Kinase Assay	Promega #V6930
DTT (Dithiothreitol), 1M, optional	
Microplate reader capable of reading luminescence	
Adjustable micropipettor and sterile tips	
30°C incubator	



Storage Conditions



This assay kit will perform optimally for up to **6 months** from date of receipt when the materials are stored as directed.

Safety



This product is for research purposes only and not for human or therapeutic use. This product should be considered hazardous and is harmful by inhalation, in contact with skin, eyes, clothing, and if swallowed. If contact occurs, wash thoroughly.

Assay Principle

The ADP-Glo™ Kinase Assay (Promega #V6930) quantifies the amount of ADP produced by a kinase upon phosphorylation of a substrate. First, addition of the ADP-Glo™ reagent terminates the reaction and quenches the remaining ATP. Second, the addition of the Kinase Detection reagent converts the produced ADP to ATP. The newly generated ATP is quantified by a luciferase reaction. The luminescent signal correlates with the amount of ADP generated by the kinase and is linear to 1 mM ATP.

Contraindications

The final concentration of DMSO in the assay should not exceed 1%.

Assay Protocol

- All samples and controls should be tested in duplicate.
- The assay should include "Blank", "Positive Control" and "Test Inhibitor" conditions.
- We recommend maintaining the diluted protein on ice during use.
- For detailed information on protein handling please refer to Protein FAQs (bpsbioscience.com).
- We recommend using XMU-MP-1 (#82586) or Staurosporine (#27002) as internal control. If not running
 a dose response curve for the control inhibitor, we recommend running the control inhibitor at 0.1X, 1X
 and 10X the IC₅₀ value shown in the validation data below.
- For instructions on how to prepare reagent dilutions please refer to Serial Dilution Protocol (bpsbioscience.com).
- 1. Thaw 5x Kinase Assay Buffer 1, 500 μM ATP, and MBP (5 mg/ml).

Optional: If desired, make 5x Kinase Assay Buffer 1 with 10 mM DTT.

2. Prepare 3 ml of **1x Kinase Assay Buffer 1** by mixing 600 μl of **5x Kinase Assay Buffer 1** with 2,400 μl of distilled water.

Note: Three (3 ml) of 1x Kinase Assay Buffer 1 is sufficient for 100 reactions.

- 3. Prepare a **Master Mix** (12.5 μ l/well): N wells x (6 μ l of 5x Kinase Assay Buffer 1 + 0.5 μ l of 500 μ M ATP + 0.5 μ l of MBP (5 mg/ml) + 5.5 μ l of distilled water).
- 4. Add 12.5 μl of Master Mix to every well.



- 5. Prepare the **Test Inhibitor** (2.5 μ l/well): for a titration prepare serial dilutions at concentrations 10-fold higher than the desired final concentrations. The final volume of the reaction is 25 μ l.
 - 5.1 If the Test Inhibitor is water-soluble: Prepare serial dilutions in 1x Kinase Assay Buffer 1, 10-fold more concentrated than the desired final concentrations.

For the positive and negative controls, use 1x Kinase Assay Buffer 1 (Diluent Solution).

OR

5.2 If the Test inhibitor is soluble in DMSO: Prepare the test inhibitor at 100-fold the highest desired concentration in 100% DMSO, then dilute the inhibitor 10-fold in 1x Kinase Assay Buffer 1 to prepare the highest concentration of the 10-fold intermediate dilutions. The concentration of DMSO is now 10%.

Prepare serial dilutions of the Test Inhibitor at 10-fold the desired final concentrations using 10% DMSO in 1x Kinase Assay Buffer 1 to keep the concentration of DMSO constant.

For positive and negative controls, prepare 10% DMSO in 1x Kinase Assay Buffer 1 (vol/vol) so that all wells contain the same amount of DMSO (Diluent Solution).

Note: The final concentration of DMSO should not exceed 1%.

- 6. Add 2.5 μl of Test Inhibitor to each well labeled "Test Inhibitor".
- 7. Add 2.5 μ l of Diluent Solution to the "Positive Control" and "Blank" wells.
- 8. Add 10 μl of 1x Kinase Assay Buffer 1 to the "Blank" wells.
- 9. Thaw **STK3 Kinase** on ice. Briefly spin the tube to recover its full content.
- 10. Dilute the protein kinase (10 μ l/well) to 2 ng/ μ l with 1x Kinase Assay Buffer 1.
- 11. Initiate the reaction by adding 10 μ l of diluted kinase to the wells designated "Positive Control" and "Test Inhibitor".

Component	Blank	Positive Control	Test Inhibitor
Master Mix	12.5 μl	12.5 μΙ	12.5 μΙ
Test Inhibitor	-	-	2.5 μΙ
Diluent Solution	2.5 μΙ	2.5 μΙ	-
1x Kinase Assay Buffer 1	10 μΙ	-	-
Diluted STK3 (2 ng/μl)	-	10 μΙ	10 μΙ
Total	25 μΙ	25 μΙ	25 μΙ

12. Incubate at 30°C for 45 minutes.



- 13. Thaw the ADP-Glo™ reagent.
- 14. At the end of the 45 minute reaction, add 25 µl of ADP-Glo™ reagent to each well.
- 15. Cover the plate with aluminum foil and incubate at Room Temperature (RT) for 45 minutes.
- 16. Thaw the Kinase Detection Reagent.
- 17. Add 50 μl of Kinase Detection reagent to each well.
- 18. Cover the plate with aluminum foil and incubate at RT for another 45 minutes.
- 19. Immediately read in a luminometer or a microplate reader capable of reading luminescence.
- 20. The "Blank" value is subtracted from all other readings.

Reading Luminescence

Luminescence is the emission of light resulting from a chemical reaction. The detection of luminescence requires no wavelength selection because the method used is emission photometry and not emission spectrophotometry.

To properly read luminescence, make sure the plate reader is set for LUMINESCENCE mode. Typical integration time is 1 second, delay after plate movement is 100 msec. Do not use a filter when measuring light emission. Typical settings for the Synergy 2 BioTek plate reader: use the "hole" position on the filter wheel; Optics position: Top; Read type: endpoint. Sensitivity may be adjusted based on the luminescence of a control assay without enzyme (typically we set this value as 100).



Example Results



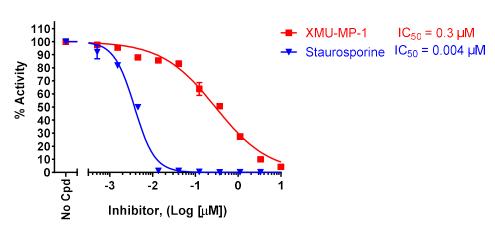


Figure 1: Inhibition of STK3 kinase activity by the inhibitors XMU-MP-1 and Staurosporine. STK3 kinase activity was measured in the presence of increasing concentrations of XMU-MP-1 (#82586) or Staurosporine (#27002). The "Blank" value was subtracted from all other values. Results are expressed as the percent of control (kinase activity in the absence of inhibitor, set at 100%).

Data shown is representative. For lot-specific information, please contact BPS Bioscience, Inc. at support@bpsbioscience.com.

Troubleshooting Guide

Visit bpsbioscience.com/assay-kits-faq for detailed troubleshooting instructions. For all further questions, please email support@bpsbioscience.com

References

Yue L., et al., 2023 International Journal of General Medicine, 16:3115-3124.

Related Products

Products	Catalog #	Size	
STK33, GST-Tag Recombinant	40093	10 μg	
LATS1, GST-Tag Recombinant	79499	10 μg	
LATS2, GST-Tag Recombinant	79042	10 μg	

Version 062024

