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Place your order with CEDARLANE® or your local distributor.

Please contact CEDARLANE® for lot specific information.

Purified Anti-Mouse CD62L No Azide, Monoclonal Antibody

CL8918NA LOT: 1821NA

DESCRIPTION:

Cedarlane's anti-mouse CD62L (L-selectin) monoclonal antibody reacts with a 90 kDa protein which is involved with the homing of lymphocytes to peripheral lymph nodes.

L-selectin is expressed on most T and B lymphocytes, neutrophils, monocytes, eosinophils.¹ Pre-incubation of lymphocytes with this antibody completely and specifically blocks binding of lymphocytes to high endothelial venules (HEV) in vitro^{2,3,6} and the migration of lymphocytes to lymph nodes in vivo.^{2,3} Polymorphonuclear cells preincubated with this antibody do not migrate to the inflammatory foci.3

Applications of CL8918NA include flow cytometry, 3,4,5 IHC4 and immunoprecipitation.3,5,6

PRESENTATION:

1 mg purified Ig buffered in PBS, no preservative, 0.2 µm filtered.

STORAGE/STABILITY:

Store at 4°C. For long term storage, aliquot and freeze unused portion at -20°C in volumes appropriate for single usage. Avoid freeze/thaw cycles. Should be handled under aseptic conditions.

For more information or to place an order please contact...



toll free: 1-800-268-5058

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5516 - 8th Line, R.R.#2, Hornby, Ontario, CANADA LOP 1E0

SPECIFICATIONS:

Clone: MEL-14

Hybridoma Production:

Immunization: Immunogen: Mouse B cell Lymphoma, 38C-14

Donor: Fischer Rat Spleen

Fusion Partner: P3 X 63Ag8.653

Specificity: Mouse CD62L (L-selectin Ly 22)

Ig Class: Rat IgG_{2a}

Format: Purified Ig buffered in PBS, no preservative. (Purifed from ascitic fluid

via Protein G Chromatography).

Antibody Concentration: 1.0 mg/ml

FLOW CYTOMETRY ANALYSIS:

Method:

- 1. Prepare a cell suspension in media A. For cell preparations, deplete the red blood cell population with Lympholyte[®]-M cell separation medium (CL5030).
- 2. Wash 2 times.
- 3. Resuspend the cells to a concentration of $2x10^7$ cells/ml in media A. Add 50 µl of this suspension to each tube (each tube will then contain $1x10^6$ cells, representing 1 test).
- 4. To each tube, add 0.1-0.2 μg* of **CL8918NA**.
- 5. Vortex the tubes to ensure thorough mixing of antibody and cells.
- 6. Incubate the tubes for 30 minutes at 4°C.
- 7. Wash 2 times at 4°C.
- Add 100 μl of secondary antibody CLCC40001 (FITC Goat anti-rat IgG (H+L)) at 1: 500 dilution.
- Incubate the tubes at 4°C for 30-60 minutes.
 (It is recommended that the tubes are protected from light since most fluorochromes are light sensitive).
- 10. Wash 2 times at 4°C in media B.
- 11. Resuspend the cell pellet in 50 µl ice cold media B.
- 12. Transfer to suitable tubes for flow cytometric analysis containing 15 μ l of propidium iodide at 0.5 mg/ml in PBS. This stains dead cells by intercalating in DNA.

Media:

- A. Phosphate buffered saline (pH 7.2) + 5% normal serum of host species + sodium azide (100 μ l of 2M sodium azide in 100 mls).
- B. Phosphate buffered saline (pH 7.2) + 0.5% Bovine serum albumin + sodium azide (100 μ l of 2M sodium azide in 100 mls).

Results:

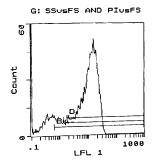
<u>Tissue Distribution by Flow Cytometry Analysis:</u>

Mouse Strain: BALB/c

Cell Concentration : $1x10^6$ cells per test Antibody Concentration Used: $0.2 \mu g/10^6$ cells

Isotypic Control: Rat IgG_{2a}

Cell Source	Percentage of cells stained above control:
Thymus	85.8%
Spleen	41.0%
Lymph Node	75.0%



Cell Source: Lymph Node
Percentage of cells stained above control: 75.0%

N.B. Appropriate control samples should always be included in any labelling studies.

* For optimal results in various applications, it is recommended that each investigator determine dilutions appropriate for individual use.

Strain Distribution by Flow Cytometry Analysis:

Procedure: see page 2

Tissue: Spleen

Cell Concentration: 1x10⁶ cells per test

Antibody Concentration Used: 0.2 µg/10⁶ cells

Strains Tested: BALB/c, C57BL/6, C3H/He, CBA/J, AKR Positive: BALB/c, C57BL/6, C3H/He, CBA/J, AKR

Negative: none

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- 3) Lewinsohn, D.M., R.F. Bargatze, E.C. Butcher 1987. Leukocyte-endothelial cell recognition: evidence of a common molecular mechanism shared by neutrophils, lymphocytes and other leukocytes. J.Immunology 138:4313-4321
- 4) Reichert, R., M. Gallitin, E. Butcher, *et al.*. 1984. A homing receptor-bearing cortical thymocyte subset: Implications for thymus cell migration and the nature of cortisone-resisatant thymocytes. Cell 38: 89-99
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