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SZABO-SCANDIC HandelsgmbH

Quellenstraße 110, A-1100 Wien

T. +43(0)1 489 3961-0

F. +43(0)1 489 3961-7

mail@szabo-scandic.com

www.szabo-scandic.com

linkedin.com/company/szaboscandic

Nordic-MUbio - Rangeerweg 5A, 6114 BC Susteren, The Netherlands
Phone : +31 (0)6 8361 1669, E-mail: info@nordicmubio.com

Product datasheet

Mouse anti Cytokeratin 10 / Keratin K10

nordicmubio.com/products/mouse-anti-cytokeratin-10-keratin-k10/MUB0319P-CE_slash_IVD

Catalog number: **MUB0319P-CE/IVD**

Clone	RKSE60
Isotype	IgG1
Product Type	Primary Antibodies
Units	0.1 mg
Host	Mouse
Species Reactivity	Canine Human Mouse Rat Swine Zebrafish
Application	Flow Cytometry Immunocytochemistry Immunohistochemistry (frozen) Western Blotting

Background

Cytokeratins are a subfamily of intermediate filament proteins and are characterized by a remarkable biochemical diversity, represented in Human epithelial tissues by at least 20 different polypeptides. They range in molecular weight between 40 kDa and 68 kDa and isoelectric pH between 4.9 – 7.8. The individual Human Cytokeratins are numbered 1 to 20. The various epithelia in the Human body usually express Cytokeratins which are not only characteristic of the type of epithelium, but also related to the degree of matuRation or differentiation within an epithelium. Cytokeratin subtype expression patterns are used to an increasing extent in the distinction of different types of epithelial malignancies. The Cytokeratin antibodies are not only of assistance in the differential diagnosis of tumors using immunohistochemistry on tissue sections, but are also a useful tool in cytopathology and flow cytometric assays.

Source
RKSE60 is a Mouse monoclonal IgG1 antibody derived by fusion of SP2/0 Mouse myeloma cells with spleen cells from a BALB/c Mouse immunized with Cytokeratins from the Human epidermis.

Product
Each vial contains 100 ul 1 mg/ml purified monoclonal antibody in PBS containing 0.09% sodium azide.

Formulation: Each vial contains 100 ul 1 mg/ml purified monoclonal antibody in PBS containing 0.09% sodium azide.

Specificity
RKSE60 reacts exclusively with Cytokeratin 10 which is present in Keratinizing stRatified epithelia and in differentiated areas of highly differentiated squamous cell carcinomas.

Applications
RKSE60 is suitable for immunoblotting, immunocytochemistry, immunohistochemistry on frozen tissues and flow cytometry. Optimal antibody dilution should be determined by titration; recommended range is 1:100 – 1:200 for flow cytometry, and for immunohistochemistry with avidin-biotinylated Horseradish peroxidase complex (ABC) as detection reagent, and 1:100 – 1:1000 for immunoblotting applications.

Storage
The antibody is shipped at ambient temperature and may be stored at +4°C. For prolonged storage prepare appropriate aliquots and store at or below -20°C. Prior to use, an aliquot is thawed slowly in the dark at ambient temperature, spun down again and used to prepare working dilutions by adding sterile phosphate buffered saline (PBS, pH 7.2). Repeated thawing and freezing should be avoided. Working dilutions should be stored at +4°C, not refrozen, and preferably used the same day. If a slight precipitation occurs upon storage, this should be removed by centrifugation. It will not affect the performance or the concentration of the product.

Caution
When used for in vitro diagnostic purposes results must be put within the context of other diagnostic tests as well as the clinical history of the patient by a certified professional before final interpretation. Analyses performed with this antibody should be paralleled by positive and negative controls. If unexpected results are obtained which cannot be attributed to differences in laboratory procedures, please contact us. This product may contain hazardous ingredients. Please refer to the Safety Data Sheets (SDS) for additional information and proper handling procedures. Dispose product remainders according to local regulations.This datasheet is as accurate as reasonably achievable, but Exalpha Biologicals accepts no liability for any inaccuracies or omissions in this information.

References

1. Ramaekers, F. C., Puts, J. J., Moesker, O., Kant, A., Huysmans, A., Haag, D., Jap, P. H., Herman, C. J., and Vooijs, G. P. (1983). Antibodies to intermediate filament proteins in the immunohistochemical identifiCation of Human tumours: an overview, Histochem J 15, 691-713. 2. Puts, J. J., Moesker, O., Kenemans, P., Vooijs, G. P., and Ramaekers, F. C. (1985). Expression of Cytokeratins in early neoplastic epithelial lesions of the uterine cervix, Int J Gynecol Pathol 4, 300-13. 3. Broers, J. L., Carney, D. N., Klein Rot, M., Schaart, G., Lane, E. B., Vooijs, G. P., and Ramaekers, F. C. (1986). Intermediate filament proteins in classic and variant types of small cell lung carcinoma cell lines: a biochemical and immunochemical analysis using a panel of monoclonal and polyclonal antibodies, J Cell Sci 83, 37-60. 4. Ramaekers, F., Huysmans, A., Schaart, G., Moesker, O., and Vooijs, P. (1987). Tissue distribution of Keratin 7 as monitored by a monoclonal antibody, Exp Cell Res 170, 235-49. 5. Van Muijen, G. N., Warnaar, S. O., and Ponec, M. (1987). Differentiation-related changes of Cytokeratin expression in cultured Keratinocytes and in fetal, newborn, and adult epidermis, Exp Cell Res 171, 331-45. 6. Bijman, J.T., Wagener, D.J.T., van Rennes, H., Wessels, J.M.C., Ramaekers, F.C.S. and van den Broek, P. (1987). Modulation of placental alkaline phosphatase activity and Cytokeratins in Human HN-1 cells by butyRate, retinoic acid, Catecholamines and histamine, Br. J. Cancer 56, 127-32. 7. Van Erp, P.E.J., Rijzewijk, J.J., Boezeman, J.B.M., Leenders, J., de Mare, S., Schalkwijk, J., van de Kerkhof, P.C.M., Ramaekers, F.C.S. and Bauer, F.W. (1989). Flow Cytometric analysis of epidermal subpopulations from normal and psoriatic skin using monoclonal antibodies anti intermediate filaments, Am J Pathol 135, 865-70. 8. Ramaekers, F., van Niekerk, C., Poels, L., Schaafsma, E., Huijsmans, A., Robben, H., Schaart, G., and Vooijs, P. (1990). Use of monoclonal antibodies to Keratin 7 in the differential diagnosis of adenocarcinomas, Am J Pathol 136, 641-55. 9. Schaafsma, H. E., Ramaekers, F. C., van Muijen, G. N., Lane, E. B., Leigh, I. M., Robben, H., Huijsmans, A., Ooms, E. C., and Ruiter, D. J. (1990). Distribution of Cytokeratin polypeptides in Human transitional cell carcinomas, with special emphasis on changing expression patterns during tumor progression, Am J Pathol 136, 329-43. 10. Smedts, F., Ramaekers, F., Robben, H., Pruszczynski, M., van Muijen, G., Lane, B., Leigh, I., and Vooijs, P. (1990). Changing patterns of Keratin expression during progression of cervical intraepithelial neoplasia, Am J Pathol 136, 657-68. 11. Franssen, M.E.J., Boezeman, J.B.M., van de Kerkhof, P.C.M. and van Erp, P.E.J. (2003). Monitoring hyperproliferative disorders in Human skin: flow cytometry of changing Cytokeratin expression, Cytometry Part B (Clinical Cytometry) 57B, 32-39.

CE Mark

CE

Protein Reference(s)

Database Name: UniProt

Accession Number: P13645

Safety Datasheet(s) for this product:
NM_Sodium Azide