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Mouse anti Cytokeratin 17 / Keratin K17

 nordicmubio.com/products/mouse-anti-cytokeratin-17-keratin-k17/MUB0325P

Catalog number: **MUB0325P**

Clone	E3
Isotype	IgG2b
Product Type	Primary Antibodies
Units	0.1 mg
Host	Mouse
Species Reactivity	Human Rat
Application	Flow Cytometry Immunocytochemistry Immunohistochemistry (frozen) Immunohistochemistry (paraffin) Western Blotting

Background

Cytokeratins are a subfamily of intermediate filament proteins and are characterized by a remarkable biochemical diversity, represented in Human epithelial tissues by at least 20 different polypeptides. They range in molecular weight between 40 kDa and 68 kDa and isoelectric pH between 4.9 – 7.8. The individual Human Cytokeratins are numbered 1 to 20. The various epithelia in the Human body usually express Cytokeratins which are not only characteristic of the type of epithelium, but also related to the degree of maturation or differentiation within an epithelium. Cytokeratin subtype expression patterns are used to an increasing extent in the distinction of different types of epithelial malignancies. The Cytokeratin antibodies are not only of assistance in the differential diagnosis of tumors using immunohistochemistry on tissue sections, but are also a useful tool in cytopathology and flow cytometric assays.

Source

E3 is a Mouse monoclonal IgG2b antibody derived by fusion of X63 Ag 8.653 Mouse myeloma cells with spleen cells from a Balb/c Mouse immunized with a cytoskeletal preparation from Rat colon.

Product

Each vial contains 100 ul 1 mg/ml purified monoclonal antibody in PBS containing 0.09% sodium azide.

Formulation: Each vial contains 100 ul 1 mg/ml purified monoclonal antibody in PBS containing 0.09% sodium azide.

Specificity

E3 reacts with Cytokeratin 17 in basal layers of pseudo-stRatified and transitional epithelia.

Applications

E3 is suitable for immunoblotting, immunocytochemistry, immunohistochemistry on frozen and paraffin-embedded tissues and flow cytometry. Optimal antibody dilution should be determined by titration; recommended range is 1:25 – 1:200 for flow cytometry, and for immunohistochemistry with avidin-biotinylated Horseradish peroxidase complex (ABC) as detection reagent, and 1:100 – 1:1000 for immunoblotting applications.

Storage

The antibody is shipped at ambient temperature and may be stored at +4°C. For prolonged storage prepare appropriate aliquots and store at or below -20°C. Prior to use, an aliquot is thawed slowly in the dark at ambient temperature, spun down again and used to prepare working dilutions by adding sterile phosphate buffered saline (PBS, pH 7.2). Repeated thawing and freezing should be avoided. Working dilutions should be stored at +4°C, not refrozen, and preferably used the same day. If a slight precipitation occurs upon storage, this should be removed by centrifugation. It will not affect the performance or the concentration of the product.

Caution

This product is intended FOR RESEARCH USE ONLY, and FOR TESTS IN VITRO, not for use in diagnostic or therapeutic procedures involving humans or animals. It may contain hazardous ingredients. Please refer to the Safety Data Sheets (SDS) for additional information and proper handling procedures. Dispose product remainders according to local regulations. This datasheet is as accurate as reasonably achievable, but Exalpha Biologicals accepts no liability for any inaccuracies or omissions in this information.

References

1. Guelstein, V. I., Tchypysheva, T. A., Ermilova, V. D., Litvinova, L. V., Troyanovsky, S. M., Bannikov, G. A. (1988). Monoclonal antibody mapping of Keratins 8 and 17 and of vimentin in normal Human mammary gland, benign tumors, dysplasias and breast

cancer, Int J Cancer 42, 147-53. 2. Troyanovsky, S. M., Guelstein, V. I., Tchipysheva, T. A., Krutovskikh, V. A., Bannikov, G. A. (1989). Patterns of expression of Keratin 17 in Human epithelia: dependency on cell position, J Cell Sci 93, 419-26. 3. Wetzel, R. H., Kuijpers, H. J., Lane, E. B., Leigh, I. M., Troyanovsky, S. M., Holland, R., van Haelst, U. J., and Ramaekers, F. C. (1991). Basal cell-specific and hyperproliferation-related Keratins in Human breast cancer. Am J Pathol 138, 751-63. 4. Moll, I., Moll, R. (1991). Comparative Cytokeratin analysis of sweat gland ducts and eccrine poromas. Arch Dermatol Res. 283, 300-09. 5. De Jong, E., van Vlijmen, I., van Erp, P., Ramaekers, F., Troyanovsky, S., Van de Kerkhof, P. (1991). Monoclonal anti-Keratin 17: A useful marker for anti-psoriatic therapies. Arch Dermatol Res 283, 480-82. 6. Smedts, F., Ramaekers, F., Troyanovsky, S., Pruszczynski, M., Link, M., Lane, B., Leigh, I., and Vooijs, P. (1992). Keratin expression in cervical cancer, Am J Pathol 141, 497-511. 7. Wetzel, R. H., Schaafsma, H. E., Leigh, I. M., Lane, E. B., Troyanovsky, S. M., Wagenaar, S. S., Vooijs, G. P., and Ramaekers, F. C. (1992). Laminin and type VII collagen distribution in different types of Human lung carcinoma: correlation with expression of Keratins 14, 16, 17 and 18. Histopathology 20, 295-303. 8. Smedts, F., Ramaekers, F., Troyanovsky, S., Pruszczynski, M., Robben, H., Lane, B., Leigh, I., Plantema, F., and Vooijs, P. (1992). Basal-cell Keratins in cervical reserve cells and a comparison to their expression in cervical intraepithelial neoplasia. Am J Pathol 140, 601-12. 9. Smedts, F., Ramaekers, F., Link, M., Laueroval, L., Troyanovsky, S., Schijf, C., and Vooijs, G. P. (1994). Detection of Keratin subtypes in routinely processed cervical tissue: implications for tumour classification and the study of cervix cancer aetiology. Virchows Arch 425, 145-155. 10. Demirkesen, C., Hoede, N., Moll, R. (1995). Epithelial markers and differentiation in adnexal neoplasms of the skin: an immunohistochemical study including individual Cytokeratins. J Cutan Pathol 22, 518-35. 11. Moll, R., et al. (1995). Differenzierungsmarker bei gynäkologischen Tumoren: Methodische und diagnostische Aspekte. In: Aktuelle Aspekte der Tumorimmunologie in der Gynäkologie. W Zuckschwerdt Verlag. 12. Litvinov, S. V., van Driel, W., van Rhijn, C. M., Bakker, H. A., van Krieken, H., Fleuren, G. J., and Warnaar, S. O. (1996). Expression of Ep-CAM in cervical squamous epithelia correlates with an increased proliferation and the disappearance of markers for terminal differentiation. Am J Pathol 148, 865-75.

Protein Reference(s)

Database Name: UniProt

Accession Number: Q04695

Safety Datasheet(s) for this product:

NM_Sodium Azide