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## Produktinformation



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Diagnostik & molekulare Diagnostik



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### SZABO-SCANDIC HandelsgmbH

Quellenstraße 110, A-1100 Wien

T. +43(0)1 489 3961-0

F. +43(0)1 489 3961-7

[mail@szabo-scandic.com](mailto:mail@szabo-scandic.com)

[www.szabo-scandic.com](http://www.szabo-scandic.com)

[linkedin.com/company/szaboscandic](https://www.linkedin.com/company/szaboscandic) 



**savyonDIAGNOSTICS**

*member of the gamida diagnostics division*

## savvy•gen GI- Rotavirus

**REF: 608-01**

**Test kit for 48 determinations**

**Store at 2-37°C**

**For Professional Use Only**  **CE**



**Savyon® Diagnostics Ltd.**

3 Habosem St. Ashdod 7761003  
ISRAEL

Tel.: +(972).8.8562920

Fax: +(972).8.8523176

E-mail: [support@savyondiagnosics.com](mailto:support@savyondiagnosics.com)



**European Authorized  
Representative: Obelis s.a.**

Boulevard Général Wahis 53  
1030 Brussels, BELGIUM

Tel: +(32) 2. 732.59.54

Fax: +(32) 2.732.60.03

E-Mail : [mail@obelis.net](mailto:mail@obelis.net)



## Intended Use

The Savvygen™GI-Rotavirus test allows the qualitative detection of Rotavirus A by real time RT-PCR in human feces. The product is intended for use in the diagnosis of Rotavirus gastrointestinal infections alongside clinical data of the patient and other laboratory tests outcomes.

## Background

Rotaviruses are triple-layered viruses that belong to the *Reoviridae* family and contain a genome of 11 segments of double-stranded (ds) RNA that encode six structural viral proteins (VP1–VP4, VP6, and VP7) and six nonstructural proteins (NSP1–NSP6). The genome is contained in the core with three proteins VP1, VP2 and VP3. Surrounding this core, there is an inner capsid composed for the main structural protein VP6 which determines the antigen classification of virus in seven groups (A to G).

Although Rotavirus serogroups A-C are human pathogens, Group A rotavirus (RVA) infection is the leading cause of acute gastroenteritis (AGE) in children aged <5 y worldwide and is associated with 500,000–600,000 deaths annually, predominantly in developing countries. Rotavirus is transmitted from person-to-person by fecal-oral route due to food, objects or water contaminated. The typical symptoms caused by these viruses are vomiting, watery diarrhea and abdominal pain.

Due to the highly conserved sequences of the *NSP3* gene, this gene has been shown to be the best target for detection of a wide variety of RVA genotypes.

## Principles of the Procedure

Savvygen™GI- Rotavirus test is designed for the identification of Rotavirus in human feces specimens to aid in the assessment of infections caused by this virus.


Savvygen™GI- Rotavirus test is based on the real time amplification of specific conserved fragments of the *NSP3* gene encoded by the Rotavirus genome. The viral RNA extracted is transcribed into cDNA using a specific primer by reverse transcription step followed immediately in the same well by polymerase chain reaction. The presence of Rotavirus A is detected by an increase in observed fluorescence during the reaction upon hydrolysis of the fluorescent probe.

The assay is based on 5' nuclease chemistry which utilizes two primers and a hydrolysis fluorogenic probe to detect the accumulation of amplified target sequence during the PCR reaction. When the polymerase begins to extend the primers, the probe is hydrolyzed by its 5' to 3' exonuclease activity causing the spatial separation of reporter and quencher. The resulting increase in fluorescence signal is proportional to the amount of amplified product in the sample and detected by the real-time PCR instrument.

is a ready-to used test which contains in each well all the necessary reagents for real time PCR assay in a stabilized format. In addition, an internal control allows the detection of a possible reaction inhibition. The amplification of the target sequence is detected through the FAM channel whereas the internal control (IC) in HEX channel.



## Materials/ Reagents Provided

| Product Description  | Contents  |
|--|---|
| <b>Savvygen™ GI -Rotavirus</b><br><b>48 reactions. Cat.# 608-01</b><br> | 6x Savvygen™ GI- Rotavirus strips<br>1x Rotavirus Positive Control<br>1x Water RNase/DNase free 1mL<br>1x Rehydration Buffer 1.8 mL<br>1x Negative Control 1 mL<br>Optical caps |

### Additional Equipment and Material Required

- Real Time PCR instrument (to check compatibility see Appendix I).
- RNA extraction kit.
- Centrifuge for 1.5 mL tube.
- Vortex.
- Micropipettes (0.5-20 µL, 20-200 µL).
- Filter tips.
- Powder-free disposal gloves

## Transport and Storage

The reagents and the test can be shipped and stored at 2-37°C until expiration date stated in the label.

The re-suspended positive control should be stored at -20°C. In order to avoid repeated freeze/thaw cycles, we recommend separating into aliquots.

Keep all reagents in the dark.

## Precautions

- This product is reserved exclusively for *in vitro* diagnostic purposes.
- Do not use after expiration date.
- The laboratory process must be one-directional, it should begin in the Extraction Area and then move to the Amplification and Detection Areas. Do not return samples, equipment and reagents to the area in which the previous step was performed.
- Follow Good Laboratory Practices. Wear protective clothing, use disposal gloves, goggles and mask.
- Do not eat, drink or smoke in areas when samples or test reagents are being used. Once you finish the test wash your hands.
- Specimens must be treated as potentially infectious as well as all reagents and materials that have been exposed to the samples and handled in the same manner as an infectious agent. Take necessary precautions during the collection, storage, treatment and disposal of samples.
- Regular decontamination of commonly used equipment is recommended, especially micropipettes and work surfaces.



## Test Procedure

### **Specimen Collection, Processing and RNA Extraction**

Stool samples should be collected in clean containers and processed as soon as possible to guarantee the quality of the test. However, the samples can be frozen at -20°C for conservation. Ensure only the amount needed is thawed because of freezing and thawing cycles are not recommended.

For pretreatment and nucleic acid isolation, it is recommended to use your optimized manual or automatic system, and even any commercially available RNA extraction kit according to manufacturer's protocol. The assay has been validated with the following extraction kits:

- Ultraclean® Tissue & Cells DNA Isolation Kit (Mobio).
- NucleoSpin® RNA virus (Machery Nagel).
- NucliSENS® EasyMAG™ platform (bioMérieux).
- QIAamp MinElute Virus Spin Kit (QIAGEN).
- QIAamp Viral RNA Mini Kit (QIAGEN).
- Invisorb® Spin Universal Kit (Strattec).
- RIDA® Xtract (R-biopharm).
- Maxwell® RSC Blood DNA Kit using the Maxwell® 16 instrument (Promega).

### **Positive Control Preparation**

Reconstitute the lyophilized *Rotavirus* Positive Control (tube of red cap) with the 100 µL of Water RNase/DNase free (tube of white cap) supplied. To ensure a complete resuspension, vortex the tube thoroughly. After first use, dispense into aliquots in order to avoid multiple freeze-thaw cycles, and store them at -20°C.

This component contains high copies number template and is a very significant contamination risk. Therefore, we recommend open and manipulate it in a separate laboratory area away from the other components.

## PCR Protocol

### **Thermo-cycler program**

Calculate the number of required reactions including samples and controls (At least one positive and one negative control). Set your thermocycler following the conditions below:

**Table 1. Real time RT-PCR conditions**

| Step                                   | Temperature | Time    | Cycles |
|--|-------------|---------|--------|
| Reverse transcription                  | 45°C        | 15 min  | 1      |
| Initial denaturation                   | 95°C        | 2 min   | 1      |
| Denaturation                           | 95°C        | 10 sec. | 45     |
| Annealing/Extension (Data collection*) | 60°C        | 50 sec. |        |

Set the fluorescence data collection during the extension step (\*) through the FAM (Rotavirus) and HEX, JOE or VIC channels (Internal Control (IC)). If you use the Applied Biosystems 7500 Fast Real-Time PCR System, the Applied Biosystems StepOne™ Real-Time PCR System or the Stratagene Mx3005P™ Real Time PCR System check that passive reference option ROX is none



**a) Reconstitute the reaction mixture of the required wells.**

Separate the number of required reactions including samples and controls. Remember that one positive and one negative control must be included in each run. Peel off protective aluminum seal from the strips/plate. Pipette 15  $\mu$ L of Rehydration Buffer (tube of blue cap) into each well.

**b) Adding samples and controls according to real-time PCR experimental plate set up.**

Pipette 5  $\mu$ L of Negative Control (tube of amber cap) into each negative control well. Pipette 5  $\mu$ L of RNA sample into each sample well. Pipette 5  $\mu$ L of reconstituted *Rotavirus* Positive Control (tube of red cap) into each positive control well. Cover the wells with the caps provided. Spin down briefly.

**c) Performing PCR.**

Place the strips/plate in the Real Time PCR instrument. Start the run.

## **Analysis and Interpretation of results**

The analysis of the results is done by the software itself of the used real time PCR system following manufacturer's instructions.

**Positive control-** The positive controls used in each run, must show an amplification curve for Rotavirus , which validates the reaction.

**Negative control-** The negative controls included in each run, must show the absence of signal for Rotavirus, which validates the reaction.

**Internal control-** The Internal Controls must show amplification curves, which verify the correct functioning of the amplification mix. Sometimes, the detection of internal control is not necessary because a high copy number of the pathogen RNA template can cause preferential amplification of target sequence.

**Positive sample-** A sample is assigned as positive for the target if the Ct value fall below 40. The internal control usually shows an amplification signal, although it might be dispensable if the amplification of the target sequence from a high copy number of RNA template can cause competition in the reaction.

**Negative sample-** A sample is assigned as negative for the target if there is no evidence of amplification signal in the detection system but the internal control is positive.

**Invalid run-** The assay should be considered as invalid and a new run should be performed if there is signal of amplification in negative control or absence of signal in the positive well.

**Note:** *If a negative sample do not show an amplification curve for the internal control, they should be retested by dilution of the original sample 1:10 or the nucleic acid extraction has to be repeated due to possible problems caused by PCR inhibitors*

**The result interpretation is summarized in table 2:**



**Table 2. Results interpretation**

| Rotavirus | Internal control  | Negative control | Positive control | Interpretation     |
|-----------|-------------------|------------------|------------------|--------------------|
| Positive  | Positive/Negative | Negative         | Positive         | Rotavirus Positive |
| Negative  | Positive          | Negative         | Positive         | Rotavirus Negative |
| Positive  | Positive          | Positive         | Positive         | Experiment fail    |
| Negative  | Negative          | Negative         | Negative         | Experiment fail    |

**Positive:** Amplification signal; **Negative:** No amplification signal

## Limitations of the test

- This test provides a presumptive diagnosis of Rotavirus A infection. All results must be interpreted together with other clinical information and laboratory findings available to the physician.
- This assay should be used only with samples from human feces. The use of other samples has not been established.
- The quality of the test depends on the quality of the sample; proper RNA from fecal specimens must be extracted. Unsuitable collection, storage and/or transport of specimens may give false negative results.
- Extremely low levels of target below the limit of detection may be detected, but results may not be reproducible.
- There is a possibility of false positive results due to cross-contamination by Rotavirus, either samples containing high concentrations of target RNA or contamination due to PCR products from previous reactions.

## Quality Control

In order to confirm the appropriate performance of the molecular diagnostic technique, an Internal Control (IC) is included in each reaction. Besides, a positive and a negative control must be included in each assay to interpret the results correctly.

## Performance Characteristics

### Clinical sensitivity and specificity

Overall, 135 fecal samples from symptomatic patients were tested by Real Time PCR using: i) Savvygen™ GI-Rotavirus test; ii) RIDA®GENE Rotavirus/Adenovirus Duplex; ii) RIDA®GENE Viral Stool Panel II (r-Biopharm) and iii) Rotavirus/Norovirus/Astrovirus Real-TM (Sacace). Rotavirus was detected in 34 samples by Savvygen™GI- Rotavirus test. Among these, only 1 sample was not able to be detected by the other tests. The results show a high sensitivity and specificity to detect Rotavirus using Savvygen™GI- Rotavirus test.

### Analytical sensitivity

This assay has a detection limit of  $\geq 10$  viral RNA copies per reaction (Figure 1).

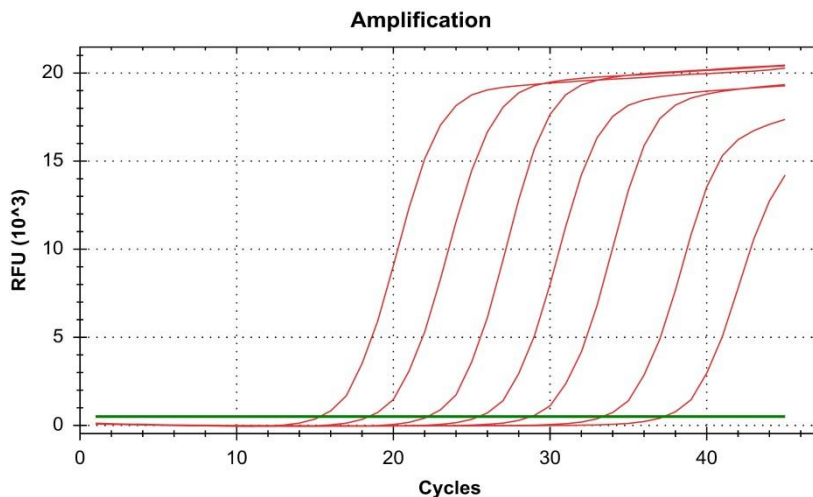


Figure 1. Amplification plot for 10-fold dilution series of Rotavirus template ranging from  $10^7$  to  $10^1$  copies/rxn

### **Analytical specificity**

The analytical specificity for Rotavirus was tested within the panel of following microorganisms, where no cross-reactivity was seen between any of the species.

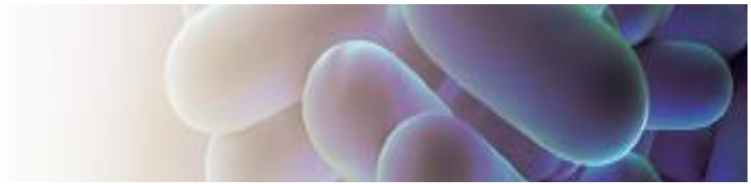
**Table 3. Cross-reactivity testing.**

|                                  |  |                                    |
|----------------------------------|--|------------------------------------|
| Adenovirus 40/41                 | <i>Cryptosporidium parvum</i>          | <i>Salmonella paratyphi A</i>      |
| Astrovirus Genotype I-VIII       | <i>Entamoeba histolytica</i>           | <i>Salmonella paratyphi B</i>      |
| Norovirus GI and GII             | <i>Enterococcus faecalis</i>           | <i>Salmonella typhimurium</i>      |
| <i>Aeromonas hydrophila</i>      | Enterotoxigenic <i>E. coli</i> (ETEC)  | <i>Salmonella bongori</i>          |
| <i>Arcobacter butzleri</i>       | Enteropathogenic <i>E. coli</i> (EPEC) | <i>Salmonella enteritidis</i>      |
| <i>Bacteroides fragilis</i>      | <i>Giardia intestinalis</i>            | <i>Salmonella enterica</i>         |
| <i>Campylobacter lari</i>        | <i>Helicobacter pylori</i>             | <i>Salmonella pullorum</i>         |
| <i>Campylobacter fetus</i>       | <i>Helicobacter hepaticus</i>          | <i>Salmonella gallinarum</i>       |
| <i>Campylobacter coli</i>        | <i>Helicobacter cinaedi</i>            | <i>Serratia liquefaciens</i>       |
| <i>Campylobacter jejuni</i>      | <i>Helicobacter heilmannii</i>         | <i>Shigella flexneri</i>           |
| <i>Campylobacter upsaliensis</i> | <i>Klebsiella oxytoca</i>              | <i>Shigella dysenteriae</i>        |
| <i>Candida albicans</i>          | <i>Listeria monocytogenes</i>          | <i>Staphylococcus aureus</i>       |
| <i>Citrobacter freundii</i>      | <i>Pseudomonas aeruginosa</i>          | <i>Vibrio parahaemolyticus</i>     |
| <i>Clostridium difficile</i>     | <i>Proteus vulgaris</i>                | <i>Y. enterocolitica</i> O:3 / O:9 |
| <i>Clostridium perfringens</i>   | <i>Salmonella typhi</i>                |                                    |

### **Analytical reactivity**

The reactivity of Savvygen™ GI-Rotavirus test was confirmed by the real time amplification using human Rotavirus A strain ATCC® VR-2018™ and Rotavirus genotypes G1P[8], G2P[4], G2P[8], G3P[8], G4P[8], G1+G3P[8], G6P[8], G9P[8], G10+G12P[8], G12P[8] and G-UDP[8] strains as template.





## **Appendix A: Compatibility of the Savvygen GI Assays with Commercial Real-Time instruments**

Savvygen™GI- Rotavirus assay has been validated on the following equipments: Applied Biosystems 7500 Fast Real-Time PCR System, Applied Biosystems StepOne™ Real-Time PCR System, Bio-Rad CFX96 Touch™ Real-Time PCR Detection System, AriaMx Real-Time PCR System, DNA-Technology DTPRime Real Time Detection Thermal Cycler. When using the Applied Biosystems 7500 Fast with strips it is recommend to place a plate holder to reduce the risk of crushed tube (Ref. PN 4388506). Additional compatible thermocyclers are listed below:

### ***Applied Biosystems***

- 7500 Fast Real-Time PCR System
- 7500 Fast Dx Real-Time PCR System
- QuantStudio™ 12K Flex 96-well Fast
- QuantStudio™ 6 Flex 96-well Fast
- QuantStudio™ 7 Flex 96-well Fast
- QuantStudio™ 3 Real-Time PCR System
- QuantStudio™ 5 Real-Time PCR System
- StepOne Plus™ Real-Time PCR System
- StepOne™ Real-Time PCR System
- ViiA™ 7 Fast Real-Time PCR System

### ***Bio-Rad***

- CFX96 Touch™ Real-Time PCR Detection System
- Mini Opticon™ Real-Time PCR Detection System

### ***Roche***

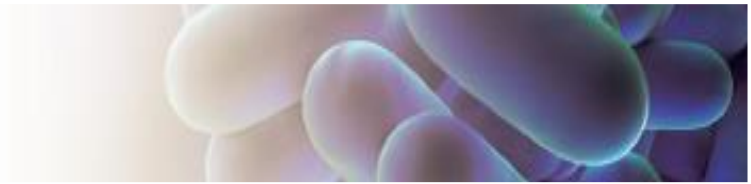
- LightCycler®480 Real-Time PCR System
- LightCycler®96 Real-Time PCR System

### ***Agilent Technologies***

- AriaMx Real-Time PCR System

### ***DNA-Technology***











- DTLite Real-Time PCR System
- DT prime Real-Time Detection Thermal Cycler



## Bibliography

1. Gautam R, Esona MD, Mijatovic-Rustempasic S, Ian Tam K, Gentsch JR, Bowen MD. Real-time RT-PCR assays to differentiate wild-type group A rotavirus strains from Rotarix(®) and RotaTeq(®) vaccine strains in stool samples. *Hum Vaccin Immunother* 2014; 10(3): 767-77.
2. Mijatovic-Rustempasic S, Tam KI, Kerin TK, Lewis JM, Gautam R, Quaye O, Gentsch JR, Bowen MD. Sensitive and specific quantitative detection of rotavirus A by one-step real-time reverse transcription-PCR assay without antecedent double-stranded-RNA denaturation. *J Clin Microbiol* 2013; 51(9): 3047-3054.
3. López L P, Carolina Cáceres D, López Medina E. Enfermedad por rotavirus. Características epidemiológicas, clínicas, prevención y manejo. *CCAP* 6(2): 45-55.

## Symbols for IVD Components and Reagents

|   |  |  |   |  |
|---|--|--|---|--|
|  For <i>in vitro</i> diagnostic use only |  Keep dry               |  Use by                           |  Manufacturer                |  Lot number       |
|  Consult instructions for use            |  Temperature limitation |  Contains sufficient for <n> test |  DIL Buffer (sample diluent) |  Catalogue number |

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