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Produktinformation

- Forschungsprodukte & Biochemikalien
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- Diagnostik & molekulare Diagnostik
- Laborgeräte & Service

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See the following pages for more information!

Lieferung & Zahlungsart

siehe unsere Liefer- und Versandbedingungen

Zuschläge

- Mindermengenzuschlag
- Trockeneiszuschlag
- Gefahrgutzuschlag
- Expressversand

SZABO-SCANDIC HandelsgmbH

Quellenstraße 110, A-1100 Wien

T. +43(0)1 489 3961-0

F. +43(0)1 489 3961-7

mail@szabo-scandic.com

www.szabo-scandic.com

linkedin.com/company/szaboscandic

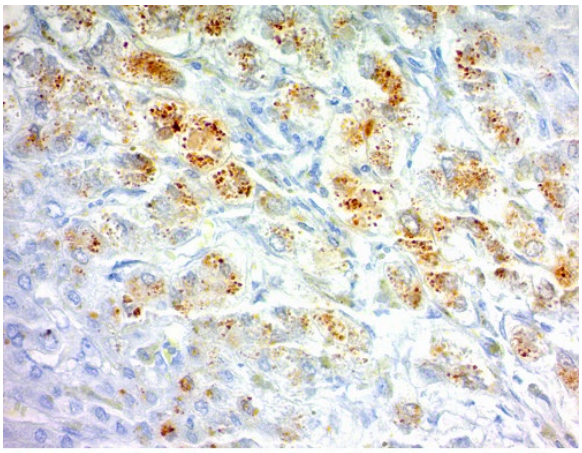
ScyTek LABORATORIES INC.

Instructions For Use

CSK-IFU

205 South 600 West Logan, Utah 84323, U.S.A. – Tel. (800) 729-8350 – Tel. (435) 755-9848 – Fax (435) 755-0015 – www.scytek.com Rev. 3, 7/19/2022

Copper Stain Kit (For Microwave)



Copper deposits in Human Liver stained with Copper Stain Kit. Viewed at 400X magnification.

Description and Principle
The Copper Stain Kit (For Microwave) is intended for the demonstration of copper deposits in tissue sections.
Rhodanine stains copper deposits in the cytoplasm of cells that are bound to copper-associated protein (CAP). Excess deposition of copper is associated with several disease states including Wilsons disease, chronic biliary obstruction, and chronic hepatitis.

Expected Results

Copper Deposits:	Light Brown to Red
Nuclei:	Blue

Kit Contents	**Storage**
1. Rhodanine Solution (Stock)	2-8°C
2. Acetate Buffer Solution, pH 8.0	18-25°C
3. Hematoxylin, Mayer's (Lillie's Mod.)	18-25°C

Suggested Controls *(not provided)*
Fetal Liver or a known positive.

Uses/Limitations
For In-Vitro Diagnostic use only.
Do not use past expiration date.
Use caution when handling reagents.
Non-Sterile
Intended for FFPE sections cut at 5-10µm.
This procedure has not been optimized for frozen sections.
Frozen sections may require protocol modification.

Storage
Mixed storage conditions. Store according to individual label instructions.

Safety and Precautions
Please see current Safety Data Sheets (SDS) for this product and components GHS classification, pictograms, and full hazard/precautionary statements.

Procedure (Standard)
Prepare **Working Rhodanine Solution** by mixing:

4 ml Rhodanine Solution (Stock) Shake Stock Solution immediately before adding to Acetate Buffer.
46 ml Acetate Buffer Solution, pH 8.0

1. Deparaffinize sections if necessary and hydrate to distilled water.

2. Place loosely capped staining jar containing Working Rhodanine in microwave and heat solution until warm but not hot.

3. Place slide in warmed Working Rhodanine Solution and microwave at full power until solution is hot. Do not allow solution to boil.

4. Cap container, gently agitate to mix evenly, and allow solution to cool on countertop to room temperature with occasional agitation.

5. Examine slide microscopically and repeat heating/cooling cycle (steps 3 & 4) until desired staining intensity is achieved.

6. Rinse slide in 2 changes of Acetate Buffer Solution, pH 8.0 for 1 minute each.

7. Stain tissue section with Hematoxylin, Mayer's (Lillie's Modification) for 5-10 seconds.

8. Rinse slide in 3 changes of Acetate Buffer Solution, pH 8.0 for 1 minute each.

9. Dehydrate slide in 3 changes of absolute alcohol.

10. Clear in 2 changes of xylene or xylene substitute, and mount in synthetic resin.

Procedure (Dropper)
Prepare **Working Rhodanine Solution** in 8ml dropper vial by mixing:

1 Drop Rhodanine Solution (Stock). Shake Stock Solution immediately before adding to Acetate Buffer.
11 Drops Acetate Buffer Solution, pH 8.0

1. Deparaffinize sections if necessary and hydrate to distilled water.

2. Place a 125ml beaker containing 100ml of water in microwave and heat to nearly boiling.

3. After heating water, carefully lay slide across the top of the beaker containing the hot water and apply 5 drops of Working Rhodanine solution. Rising heat and steam from water will warm slide and enhance staining.

4. Allow Working Rhodanine solution to incubate on tissue section until water has cooled to room temperature. Check occasionally to ensure that the tissue section is not allowed to dry.

5. Examine slide microscopically and repeat heating/cooling cycle (steps 2-4) until desired staining intensity is achieved.

6. Rinse slide in 5-6 drops of Acetate Buffer Solution, pH 8.0 for 1 minute, shake off excess and repeat.

7. Stain tissue section with 5-6 drops of Hematoxylin, Mayer's (Lillie's Modification) for 5-10 seconds.

8. Rinse slide in 5-6 drops of Acetate Buffer Solution, pH 8.0 for 1 minute, shake off excess and repeat twice more.

9. Dehydrate slide in 3 changes of absolute alcohol.

10. Clear in 2 changes of xylene or xylene substitute, and mount in synthetic resin.

References

1. Sheehan, DC., Hrapchak, BB. Theory and Practice of Histotechnology; 1980, page 230.
2. Lindquist, RR. Studies on the Pathogenesis of Hepatolenticular II: Cytochemical methods for the location of copper. Arch Pathol; 1969, Volume 87: page 370.

ScyTek Laboratories, Inc.
205 South 600 West
Logan, UT 84321
435-755-9848
U.S.A.

CE IVD

EC REP

Emergo Europe
Prinsessegracht 20
2514 AP The Hague, The Netherlands